# SHORT COMMUNICATION

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# Variation at 16 STR loci in Rwandans (Hutu) and implications on profile frequency estimation in Bantu-speakers

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**Abstract** A data set of 16 autosomal STRs (the 13 CODIS loci plus HumCD4, HumFES, HumF13A1) was obtained in a sample of 52 unrelated Hutus from Rwanda. Genotypes at all loci met Hardy-Weinberg expectations with the exception of HumCSF1PO. No significant evidence of association across alleles at independent loci was obtained. Statistical parameters demonstrated the forensic usefulness of the analysed systems (combined PE=0.9999996, combined PD=1:2.27 $\times$ 10<sup>18</sup>). Pairwise comparisons showed that the Hutu gene pool differs substantially from that of other Bantu-speaking populations suggesting the use of ethnic-specific population databases in forensic casework analysis. The introduction of a non-negligible bias was confirmed by calculating the differences between multiple-locus profile frequencies of western and eastern Bantoids using local and non-local reference databases.

**Keywords** STR-profiles · Hutu · Rwanda · CODIS · Bantu-speakers

## Introduction

Standard sets of short tandem repeat loci (STRs) have been adopted world-wide in routine forensic practice both for casework analyses and data banking. The 13 autosomal loci HumCSF1PO, D3S1358, D5S818, D7S820, D8S1179,

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V. Coia · G. Destro-Bisol Department of Animal and Human Biology, University of Rome "La Sapienza", P.le A. Moro, Rome, Italy D13S317, D16S539, D18S51, D21S11, HumFGA, HumTH01, HumTPOX and HumVWA, are included in the most widely used commercial kits for DNA profiling (AmpF*l*STR Profiler Plus+ COfiler and AmpFlSTR Identifiler by Applied Biosystems; GenePrint Powerplex 16 System by Promega Corporation), form the U.S. CODIS database (Combined DNA Index System), have been technically validated [1], and satisfy the recommendations of the European Network of Forensic Science Institutes (ENFSI). Further STR loci have been validated for forensic applications and included in web repositories, among others HumCD4, HumFES, HumF13A1 [2, 3].

The availability of rapid and reliable genotyping procedures have facilitated the collection of reference data for many human populations as they are needed to confidently infer statistical estimates of multiple-loci DNA profiles [4, 5]. World-wide population surveys based on microsatellite loci showed that genetic variation is highest in Africans [6, 7] and that the observed linear gradient of decreasing diversity away from Africa is free of ascertainment bias [8].

Within sub-Saharan Africa, the region surrounding Lake Victoria is one of the most populated areas. However, STR-based DNA profiles of the human groups living there are virtually missing from anthropological and forensic databases, the unique exception being the Ugandans for five loci [9]. The Hutus (11.3 million according to The World Factbook [10]) are the largest Bantu-speaking group from Rwanda and Burundi (80–90% of the whole population), where they have been living for approximately 2,000 years as farmers. Even though the life expectancy rate at birth is the lowest in the world (39.3 years) and they have played the major role in terms of victims (around 1 million) and refugees (around 2 million) during the civil war of 1994, the population size has tripled in the last two generations [10, 11].

In this paper, we present the allele frequencies at 16 autosomal STR loci (those included in the CODIS database plus HumCD4, HumFES, HumF13A1) in a population sample of 52 Hutus from north-eastern Rwanda, compare them with those of other sub-Saharian populations with the

same linguistic affiliation (Niger-Kordofan/Benue-Congo/ Bantoid), and estimate multi-locus profile frequency shifts using local and non-local reference databases. We aimed at enlarging the panel of STR reference data for African populations and at evaluating its use for forensic purposes.

#### Materials and methods

The samples were collected among the workers of the catholic parish at Nyarurema (1°21'S, 30°19'E), in north-eastern Rwanda. The study has been performed in accordance with the ethical standards, all donors gave their informed consent prior to being included in the research and personal data have been treated anonymously.

From 3 to 6 eyebrows were pulled out and stored in eppendorf tubes for 5–7 days until their arrival at the laboratory of Molecular Anthropology of the University of Pisa. DNA was isolated from hair bulbs following the method of Higuchi et al. [12]. The genotyping of the CODIS core loci (HumCSF1PO, D3S1358, D5S818, D7S820, D8 S1179, D13S317, D16S539, D18S51, D21S11, HumFGA, HumTH01, HumTPOX, HumVWA) plus the Amelogenin locus for sex typing was carried out in the Laboratory of Forensic Haematology of the Catholic University of Rome, by AmpF*l*STR Profiler Plus and AmpF*l*STR COfiler PCR amplification kits (Applied Biosystems) according to product instructions. Fragment sizes were detected by the ABI PRISM 310 Genetic Analyzer (Applied Biosystems) using Genescan 500 ROX as internal size standard and sequenced allele ladders. The genotyping of HumCD4, HumFES and HumF13A1 loci was performed in the Laboratory of Molecular Anthropology of the University "La Sapienza" of Rome. PCR conditions were as previously described [13]. The amplified products were analysed in denaturing polyacrylamide gels using a semi-automated DNA sequencer (A.L.F. express, Pharmacia Biotech). Allele and internal standards were used for typing. Allele nomenclature followed the recommendations of the European DNA profiling group (EDNAP) [14].

Departures from the Hardy-Weinberg (HWE) and linkage equilibrium (LD) were estimated by exact tests using a Markov chain method (200,000 runs) with the GENEPOP 3.1 software [15]. Pairwise population differentiations were estimated by Fisher exact tests [16] with the ARLEQUIN 2.000 software [17]. The unbiased heterozygosity (H) and relative standard errors were calculated according to equations 8.4 and 8.7 in Nei [18], the polymorphism information content (PIC) as described in Botstein et al. [19] and the power of discrimination (PD) according to Weir [20].

### Results and discussion

The distributions of the allele frequencies (analytical results are available from the authors upon request) and the statistical parameters of forensic interest computed from the 16 STR systems are given in Tables 1 and 2. Although the genotyped sample is small it has to be considered as representative of the allele diversity within Hutu population. In fact, we observed worldwide very rare variant alleles (i.e. D18S51\*15.2, HumFGA\*31.2) and an average number of alleles/locus higher than those from larger African population data sets (i.e. the Bubi from Equatorial Guinea or Blacks from Zimbabwe, see Table 3). The amelogenin typing always matched the phenotypic sex of the donors [21].

Only one probability test showed significant departures from the HWE equilibrium, at the HumCSF1PO locus. The *p*-value remained under the conventional limit of significance also after the application of the Bonferroni correction for multiple tests [22]. The use of a more sensitive test (one-tail DE test, *p*=0.021) demonstrated that such deviation was due to homozygote excess.

Allele	HumCD4 $(2N=100)$	HumCSF1PO $(2N=100)$	D3S1358 $(2N=104)$	D5S818 $(2N=102)$	D7S820 $(2N=100)$	D8S1179 $(2N=104)$	D13S317 $(2N=104)$	D16S539 $(2N=100)$
3.2								
5	0.2100							
6	0.0500							
7	0.0500	0.0800			0.0300			
8	0.1000	0.0100		0.0588	0.1100		0.0481	0.0700
9	0.0200	0.1700			0.1200			0.1300
10	0.1900	0.2700		0.0294	0.4400	0.0192	0.0288	0.1300
11	0.1800	0.2200	0.0096	0.2451	0.2100	0.0577	0.2692	0.2900
12	0.0900	0.2400	0.0096	0.3824	0.0800	0.1538	0.3942	0.2300
13	0.0500	0.0100	0.0096	0.2843	0.0100	0.1538	0.1827	0.1400
14	0.0600		0.0673			0.2692	0.0769	
15			0.2692			0.2788		0.0100
16			0.3365			0.0385		
17			0.2404			0.0288		
18			0.0577					
H	$0.862 \pm 0.009$	$0.790 \pm 0.009$	$0.752 \pm 0.013$	$0.712 \pm 0.014$	$0.732 \pm 0.022$	$0.800 \pm 0.012$	$0.733 \pm 0.017$	$0.809 \pm 0.011$
<b>PIC</b>	0.8421	0.7515	0.7065	0.6551	0.6945	0.7675	0.6872	0.7773
PD	0.9641	0.9199	0.8948	0.8616	0.8923	0.9296	0.8844	0.9345
PE	0.7162	0.5758	0.5197	0.4534	0.516	0.6035	0.4992	0.6159
<b>HWE</b> Exact test	$0.108 \pm 0.006$	$0.001 \pm 0.000$	$0.740 \pm 0.008$	$0.204 \pm 0.004$	$0.689 \pm 0.007$	$0.546 \pm 0.008$	$0.292 \pm 0.006$	$0.672 \pm 0.006$

**Table 1** Allele frequencies and relevant forensic statistics computed at the 16 STR loci in the Hutu population (Rwanda)

Values under the 0.05 level are in boldtype.





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Out of 78 comparisons only 2 exact tests for independence across loci gave probability values below the 0.05 level, less than expected on the basis of a 5% level of significance (3.9 pairs of loci). Therefore, the frequency of the multiple locus DNA profile can be estimated with a certain confidence by the product rule, namely the multiplication of individual locus profile frequencies. The estimated values for combined PE (0.9999996) and combined PD  $(1: 2.27 \times 10^{18})$  demonstrate the high forensic efficiency of the investigated set of STR loci in the Hutu population. The statistical support for Hutus is somewhat comparable with the combined values obtained by the Powerplex 16 System on African-Americans (PE=0.9999996, PD=1:1.42× 1018) [23].

Linguistic and genetic evidence supports the archaeological hypothesis of a recent dispersal of Bantu farmers within the last 3000–4000 years [24, 25]. However, we observed significant differences at 11 loci between Hutu STR distributions and those of other Bantu-speakers groups (Table 3). The systems HumCSF1PO, D8S1179, HumFGA, HumTH01, HumWA and HumCD4, demonstrated to be highly discriminative as significant probability values were observed for most of the comparisons and global tests (data not shown), were always below the 0.001 significance level. This finding suggest an unfair bias (i.e. an overestimate against the suspect) in applying non-specific population databases to calculate matching probabilities when Bantu-speakers are considered as the source of DNA evidence [26].

In order to quantify the bias introduced when the Hutu database is used with non-Hutu Bantoids, we calculated the difference between 9-locus (HumCSF1PO, D3S1358, D18S51, D21S11, HumTH01, HumTPOX, HumVWA, HumFES, HumF13A1) profile frequencies of genotyped individuals belonging to the eastern (Hutus) and the western clades of Bantu dialects inferred by using local and non-local databases. To obtain the most conservative estimate of profile frequency shifts we chose the population with the least genetic differentiation from Hutus (Ewondo and Bamileke from Cameroon, see Table 3) and selected only those genotypes having no allele missing in the reference database. The number of individuals considered thus ranged from 50, comparing Ewondo genotypes against the Bamileke database, and 21, comparing Hutus genotypes against the Ewondo database (76.9% and 40.4%, respectively of the whole available data set). The presence of private alleles within each data set due to both low sample sizes and different population histories, accounts for this reduction.

When profile frequencies for Bamileke or Ewondo individuals were estimated against the other western Bantu database, average frequencies were 16 and 17 times lower, respectively than the frequencies estimated against the ethnic-specific database and the largest differences were about 2 orders of magnitude (from  $9.24 \times 10^{-12}$  to  $7.99 \times$  $10^{-14}$  and from  $1.02 \times 10^{-13}$  to  $1.19 \times 10^{-15}$ ). Conversely, the average matching probabilities for western Bantu genotypes were significantly lower when using the Hutu database and, vice versa, when using a Bamileke or Ewondo database for Hutu genotypes. In this cases they were from 36 to 358 times lower than the frequencies obtained by using population-specific databases, the largest shift exceeded 3 orders of magnitude (from  $6.34 \times 10^{-14}$  to  $2.01\times10^{-17}$ ), and often the reciprocal of the upper frequency estimate approached the size of the world population  $({\sim}7{\times}10^9)$ .

Therefore, our conclusions are different from those recently drawn for Arabic speaking populations [27]: the use of non-local reference population data in forensic casework involving Bantu speakers or, more generally, sub-Saharian Africans, may yield matching probability estimates which differ several orders of magnitude, with nonnegligible impact on the jury.

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